



## Supplementary Material

# Development and Clinical Evaluation of a Direct Amplification Method to Diagnose Canine Parvovirus and Canine Distemper Viral Infections in Dogs without Nucleic Acid Extraction

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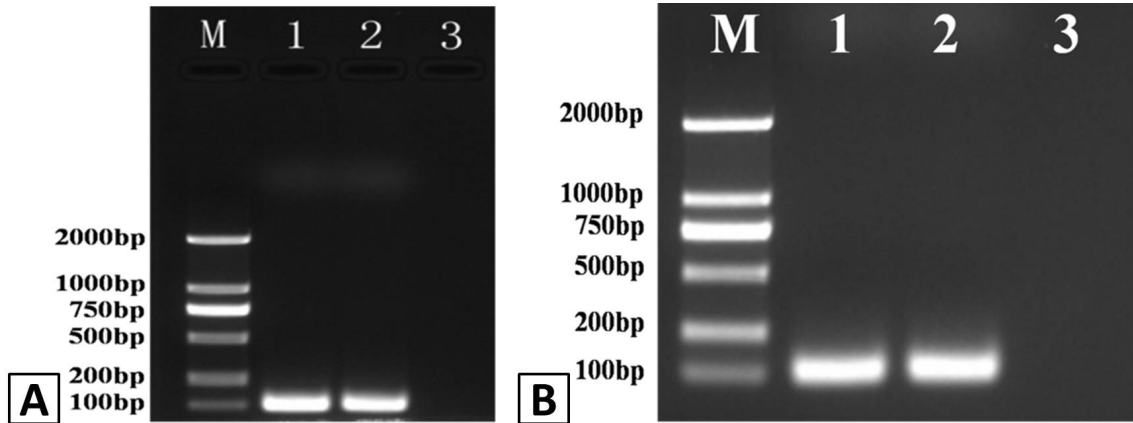
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**Supplementary Table I.- Intra-batch coefficient variant (CV) of the real-time Taq-Man PCR/RT-PCR for CPV and CDV.**

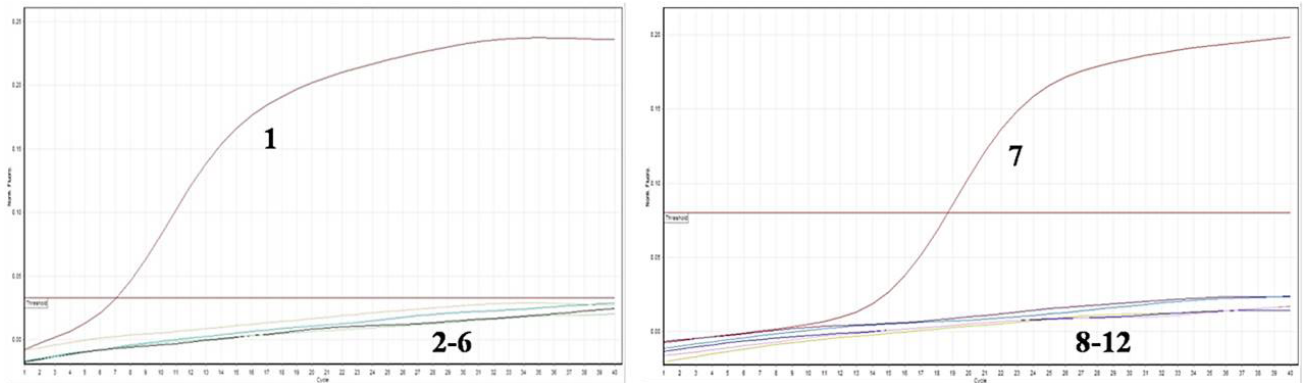
	Concentration of sample (copies· $\mu$ L <sup>-1</sup> )	C <sub>t1</sub>	C <sub>t2</sub>	C <sub>t3</sub>	Average of C <sub>t</sub>	Variance	Intra-batch CV (%)
CPV	7.44×10 <sup>7</sup>	15.82	15.88	15.50	15.73	0.042	1.30
	7.44×10 <sup>5</sup>	22.75	22.85	22.79	22.79	0.003	0.22
	7.44×10 <sup>3</sup>	29.57	29.53	29.52	29.52	0.007	0.09
CDV	4.20×10 <sup>6</sup>	13.86	14.17	13.99	14.01	0.024	1.11
	4.20×10 <sup>4</sup>	20.04	19.98	20.27	20.10	0.023	0.76
	4.20×10 <sup>2</sup>	26.22	26.14	25.91	26.09	0.026	0.62

**Supplementary Table II.- Inter-batch coefficient variant (CV) of the real-time Taq-Man PCR/RT-PCR for CPV and CDV.**

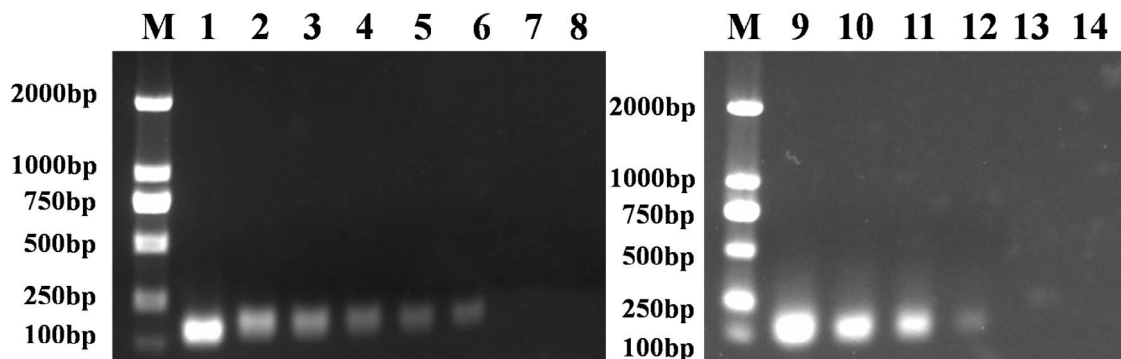
	Concentration of sample (copies· $\mu$ L <sup>-1</sup> )	C <sub>t1</sub>	C <sub>t2</sub>	C <sub>t3</sub>	Average of C <sub>t</sub>	Variance	Inter-batch CV (%)
CPV	7.44×10 <sup>7</sup>	15.88	15.97	15.68	15.84	0.022	0.94
	7.44×10 <sup>5</sup>	22.86	23.20	23.03	23.03	0.029	0.73
	7.44×10 <sup>3</sup>	29.55	29.77	29.44	29.59	0.028	0.57
CDV	4.20×10 <sup>6</sup>	13.90	14.27	14.12	14.06	0.021	1.02
	4.20×10 <sup>4</sup>	20.13	20.21	20.43	20.26	0.024	0.77
	4.20×10 <sup>2</sup>	26.11	26.27	26.04	26.14	0.014	0.45



Supplementary Fig. S1. Detection of CPV and CDV plasmid standards by PCR and RT-PCR. PCR products (111 bp and 100 bp) were visualized by gel electrophoresis. A: Lane 1, Marker; Lanes 2 and 3, CPV plasmid; Lane 4, negative control; B: Lane 1, Marker; Lanes 2 and 3, CDV plasmid; Lane 4, negative control.



Supplementary Fig. S2. Real-time PCR specificity was validated using nucleic acids from CDV, CAV-1, CAV-2, CPIV, RABV and plasmid standards for CPV; all the samples tested negative except for the CPV plasmid standard. Real-time RT-PCR specificity was validated using nucleic acids from CPV, CAV-1, CAV-2, CPIV, RABV and the plasmid standard for CDV; all the samples tested negative except for the CDV plasmid standard. 1, CPV plasmid standard; 2–6, CDV, CAV-1, CAV-2, CPIV and RABV nucleic acids; 7, CDV plasmid standard, 8–12, CPV, CAV-1, CAV-2, CPIV and RABV nucleic acids.



Supplementary Fig. S3. Sensitivity of PCR and RT-PCR for detecting CPV and CDV plasmids. The serial 10-fold plasmid dilutions (CPV:  $7.44 \times 10^8$  copies  $\cdot \mu\text{L}^{-1}$  to  $7.44 \times 10^2$  copies  $\cdot \mu\text{L}^{-1}$ , CDV:  $4.20 \times 10^7$  copies  $\cdot \mu\text{L}^{-1}$  to  $4.20 \times 10^3$  copies  $\cdot \mu\text{L}^{-1}$ ) were visualized by gel electrophoresis. A minimum of  $7.44 \times 10^3$  copies of CPV (A) and  $4.20 \times 10^4$  copies of CDV (B) were detected. M, 2K Marker; 1–7, equal dilutions of the CP.